

Reagents: i = SOCl₂, ii = R₂NH₂, iii = LAH, iv = R₂COCl

ethyl-5-oxo-3-pyrrolidinylcarboxylic acid (126 g, 0.5 mole) in CHCl₃ (200 ml) was added to freshly distd SOCl₂ (250 g, 2.1 mole). The reaction mixture was heated under reflux for 2 hr. The solvent and unreacted SOCl₂ were evapd under reduced pressure and the residual oil used without further purification.

Preparation of Amides. Method A.—Amines and *N*-phenethyl-5-oxo-3-pyrrolidinylcarbonyl chloride in equimolar quantities were allowed to react in an excess of 2% NaOH. A solid sepd on cooling, this was collected, washed (H₂O), and recrystd.

Method B.—Amines (2.0 moles) and *N*-phenethyl-5-oxo-3-pyrrolidinylcarbonyl chloride (1.0 mole) were allowed to react in dry CHCl₃ at -60°. A solid, which sepd on storage, was collected and washed (CHCl₃) and the washings were added to the filtrate. The combined washings were dried (Na₂SO₄) and evapd. The residual oils solidified on cooling and were recrystd. For details see Table II.

***N*-Phenethyl-3-pyrrolidinylmethylamines (4).**—LAH (4.7 g, 0.15 mole) was suspended in dry dioxane (150 ml) in a Soxhlet apparatus. *N*-Phenethyl-5-oxo-3-pyrrolidinylcarboxamides (0.1 mole) were packed into the thimble and extracted. The products were worked up in the usual way and purified as such or characterized as acyl derivatives.

***N*-Phenethyl-3-pyrrolidinylmethylaniline (4a)** was redistilled under reduced pressure, the fraction boiling at 190° (1.5 mm) was collected: yield 14.75 g (52%). *Anal.* (C₁₉H₂₄N₂) C, H, N.

***N*-Phenethyl-3-pyrrolidinylphenethylamine·2HBr (4b)** was recrystd from EtOH-Et₂O: mp 255-256°; yield 31.95 g (68%). *Anal.* (C₂₁H₃₀Br₂N₂) C, H, N.

***N*-(Substituted phenyl)-*N'*-(1-phenethyl-3-pyrrolidinyl)acetamides (1).** **Method A.**—*N*-Phenethyl-3-pyrrolidinylmethylamines (0.1 mole) in dry CHCl₃ (100 ml) were added to anhyd NaHCO₃ (0.15 mole). The suspension was cooled to 0° and the appropriate acid chloride (0.2 mole) added. The reaction mixture was heated under reflux for 5 hr and filtered hot. The filtrate was dried (Na₂SO₄) and evapd. The residual oils were distd under reduced pressure.

Method B.—*N*-Phenethyl-3-pyrrolidinylmethylamines (0.1 mole) in dry CHCl₃ (100 ml) were treated with the appropriate acid chloride (0.3 mole) and the reaction mixture was heated under reflux for 2 hr. Excess acid chloride and solvent was distd off under reduced pressure. The residue was basified by addn of 36% KOH and the base extd with Et₂O (4 × 25 ml). The products were isolated as in method A. Compound 4 was insol in Et₂O.

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2,2'-Dialkoxybenzhydrylamides and 2,2'-Dialkylbenzhydryl Esters of *N,N*-Disubstituted α -Amino Acids. Synthesis and Pharmacological Evaluation

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Isomers of lidocaine which do not have two Me groups ortho to the anesthesiophore group differ markedly in anesthetic properties.¹ Obviously the considerable steric hindrance plays an important role in this pharmacological activity. Moreover, the replacement of a functional NH by O can lead to isosteric compounds of similar properties.^{2, 3}

The title compounds were chosen for study because they contain a key structural feature of lidocaine: steric hindrance; moreover, we were interested in studying what effect replacement of NHCO by OCO in this type of molecules would have on the activity profile.

The synthesis of the intermediate halo esters was achieved under mildly basic conditions. Attempts to prepare them by boiling di-*o*-tolylearbinol and ClCO-CH₂Cl were unsuccessful. di-*o*-tolylechloromethane⁴ was obtained.

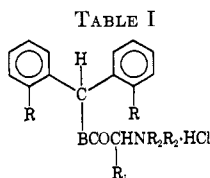
An attempt was made to correlate local anesthetic potency with the bond order of the CO linkage as measured by the CO stretching frequency. A previous correlation of this type has been reported.^{5, 6} Examination of Table I shows there is no correlation between the CO absorption frequency and the local anesthetic potency. Direct comparison of esters and amides perhaps should not be made, particularly with the hindered amides reported here, since amides in general are representatives of a lower absorption frequency.

Biological Results.—Compounds 1-28 were tested for their local anesthetic activity and the results of the observations are summarized in Table I. Potency and duration of local anesthetic activity were assessed by the Bülbring and Wajda technique.⁷ Aliquots of 0.25% solns of 1-20 in distd H₂O were injected intradermally in guinea pigs. Compounds 21-28, because of instability in H₂O, were injected at a dose level of 0.25% in propylene glycol. Local anesthesia was indicated by the absence of a flinching response when the treated site was pricked at 5, 10, 15, 30, 60, 120, and 180 min after injection. Lidocaine was used for comparison throughout the experiments.

It is apparent from these primary results that 3 is somewhat more active than lidocaine itself. However, the injection site was inflamed and edematous. Twenty-four hours after the experiment the animals

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- (1) I. Fischer and N. M. Löfgren, *Acta Chem. Scand.*, **4**, 1408 (1950).
- (2) E. Brancaccio and A. Larizza, *Farmaco Ed. Sci.*, **19**, 986 (1964).
- (3) P. Pietra, M. L. Bruschi, and F. Trimarchi, *ibid.*, **21**, 785 (1966).
- (4) M. Yasue, *J. Pharm. Soc. Jap.*, **76**, 95 (1956).
- (5) A. M. Galinsky, J. E. Gearien, A. J. Perkins, and S. V. Susina, *J. Med. Chem.*, **6**, 320 (1963).
- (6) W. O. Foye, H. B. Levine, and W. L. McKenzie, *ibid.*, **9**, 61 (1966).
- (7) E. Bülbring and I. Wajda, *J. Pharmacol. Exp. Ther.*, **85**, 78 (1945).



Compd	R	R ₁	N R ₂ R ₂	B	Yield, %	Mp, °C	Formula ^c	Carbonyl absorption frequency, cm ⁻¹	Surface local anesthetic activity ^d
1	OMe	H	-N(CH ₃) ₂	NH	91	205-206	C ₁₉ H ₂₃ ClN ₂ O ₃	1666	0.80
2	OMe	H	-N(C ₂ H ₅) ₂	NH	88	200-201	C ₂₁ H ₂₉ ClN ₂ O ₃ ^b	1672	0.96
3	OMe	H	-N(CH ₂) ₃ CH ₂	NH	85	145-146	C ₂₁ H ₂₇ ClN ₂ O ₃	1658	1.30
4	OMe	H	-N(CH ₂) ₄ CH ₂	NH	80	203-204	C ₂₂ H ₂₉ ClN ₂ O ₃	1656	0.90
5	OMe	H	-N(CH ₂) ₂ OCH ₂ CH ₂	NH	82	207-208	C ₂₁ H ₂₇ ClN ₂ O ₄	1658	0.70
6	OMe	Me	-N(CH ₃) ₂	NH	78	246-247	C ₂₀ H ₂₇ ClN ₂ O ₃	1661	<0.10
7	OMe	Me	-N(C ₂ H ₅) ₂	NH	81	229-230	C ₂₂ H ₃₁ ClN ₂ O ₃ ^d	1672	0.50
8	OMe	Me	-N(CH ₂) ₃ CH ₂	NH	79	208-209	C ₂₂ H ₂₉ ClN ₂ O ₃	1669	0.50
9	OMe	Me	-N(CH ₂) ₄ CH ₂	NH	80	220-221	C ₂₃ H ₃₁ ClN ₂ O ₃	1661	0.80
10	OMe	Me	-N(CH ₂) ₂ OCH ₂ CH ₂	NH	76	209-210	C ₂₂ H ₂₉ ClN ₂ O ₄	1669	0.80
11	OEt	H	-N(CH ₃) ₂	NH	79	199-200	C ₂₁ H ₂₉ ClN ₂ O ₃	1666	0.96
12	OEt	H	-N(C ₂ H ₅) ₂	NH	76	191-192	C ₂₃ H ₃₃ ClN ₂ O ₃	1669	0.96
13	OEt	H	-N(CH ₂) ₃ CH ₂	NH	78	193-194	C ₂₃ H ₃₁ ClN ₂ O ₃	1658	0.80
14	OEt	H	-N(CH ₂) ₄ CH ₂	NH	79	194-195	C ₂₄ H ₃₃ ClN ₂ O ₃	1667	0.40
15	OEt	H	-N(CH ₂) ₂ OCH ₂ CH ₂	NH	74	203-204	C ₂₃ H ₃₁ ClN ₂ O ₄	1656	0.80
16	OEt	Me	-N(CH ₃) ₂	NH	81	237-238	C ₂₂ H ₃₁ ClN ₂ O ₃	1667	<0.10
17	OEt	Me	-N(C ₂ H ₅) ₂	NH	78	234-235	C ₂₄ H ₃₅ ClN ₂ O ₃	1669	<0.10
18	OEt	Me	-N(CH ₂) ₃ CH ₂	NH	76	220-221	C ₂₄ H ₃₃ ClN ₂ O ₃	1672	<0.10
19	OEt	Me	-N(CH ₂) ₄ CH ₂	NH	77	246-247	C ₂₅ H ₃₅ ClN ₂ O ₃	1669	<0.10
20	OEt	Me	-N(CH ₂) ₂ OCH ₂ CH ₂	NH	75	216-217	C ₂₄ H ₃₃ ClN ₂ O ₄	1675	0.10
21	Me	H	-N(C ₂ H ₅) ₂	O	59	198-200	C ₂₁ H ₂₈ ClNO ₂	1764	0.98
22	Me	H	-N(CH ₂) ₃ CH ₂	O	44	195-196	C ₂₁ H ₂₆ ClNO ₂ ^e	1751	1.04
23	Me	H	-N(CH ₂) ₄ CH ₂	O	63	177-179	C ₂₂ H ₂₈ ClNO ₂ ^f	1748	0.72
24	Me	H	-N(CH ₂) ₂ OCH ₂ CH ₂	O	52	173-175	C ₂₁ H ₂₆ ClNO ₃	1748	0.39
25	Me	Me	-N(C ₂ H ₅) ₂	O	28	147-148	C ₂₂ H ₃₀ ClNO ₂	1730	1.02
26	Me	Me	-N(CH ₂) ₃ CH ₂	O	68	161-162	C ₂₂ H ₂₈ ClNO ₂ ^g	1742	0.86
27	Me	Me	-N(CH ₂) ₄ CH ₂	O	66	168-169	C ₂₃ H ₃₀ ClNO ₂	1730	1.02
28	Me	Me	-N(CH ₂) ₂ OCH ₂ CH ₂	O	58	135-136	C ₂₂ H ₂₈ ClNO ₃	1739	1.10

^a Values refer to lidocaine, assigned a relative anesthetic potency of 1. ^b C: calcd, 62.55; found, 63.00. ^c All compounds were analyzed for C, H, Cl, N. ^d C: calcd, 64.94; found, 64.50. ^e H: calcd, 7.23; found, 7.75. ^f C: calcd, 70.68; found, 69.90. ^g H: calcd, 7.49; found, 8.20.

were killed. Subsequent histological examination of the areas showed acute inflammatory response with proliferation of polynuclear cells. However, 7 days after the injection, tissue irritation had disappeared.

Experimental Section⁸

2,2'-Dialkoxy-N-formylbenzhydrylamines.—(NH₄)₂CO₃ (22 g) and HCOOH (22 ml) were mixed, heated cautiously, and then

(8) All melting points were taken in Fischer-John apparatus and are uncorrected. Ir spectra were measured on a Perkin-Elmer Model 137 E

slowly distd until the temp was about 160°. To the hot mixture 10 g of the appropriate 2,2'-dialkoxybenzhydrylamine⁹ was added and the temp raised to 180-185° for 10 hr. After cooling the mixture was poured into 40 ml of H₂O and the product was collected, washed, dried, and recrystd (EtOH).

2,2'-Dimethoxy-N-formylbenzhydrylamine (29) was obtained in 98% yield (10.8 g), mp 180-181°. Anal. (C₁₆H₁₇NO₃) C, H, N.

Infracord. Microanalyses were performed at this laboratory. Where analyses are indicated only by symbols of the elements, analytical results obtained for those elements were within ±0.4% of the theoretical values.

(9) C. Graebe and A. Feer, *Ber.*, **19**, 2610 (1886).

2,2'-Diethoxy-N-formylbenzhydrylamine (30) was obtained in 93% yield (10.2 g), mp 170–171°. *Anal.* (C₁₈H₂₁NO₃) C, H, N.

2,2'-Dialkoxybenzhydrylamines.—A suspension of 10 g of the appropriate N-formyl derivative in 50 ml of HCl 5% was refluxed until a clear soln was obtd. After cooling the amine·HCl crystd.

2,2'-Dimethoxybenzhydrylamine·HCl (31) was obtained in 76% yield (7.8 g), mp 246–247°. *Anal.* (C₁₅H₁₈ClNO₂) C, H, Cl, N.

2,2'-Diethoxybenzhydrylamine·HCl (32) was obtained in 85% yield (9.8 g), mp 239–240°. *Anal.* (C₁₇H₂₂ClNO₂) C, H, Cl, N.

2,2'-Dialkoxybenzhydrylamides.—A soln of 0.01 mole of Cl-COCH₂Cl or BrCOCHBrCH₃ in 10 ml of C₆H₆ was added over a period of 0.5 hr to a cold, stirred soln of 0.02 mole of the appropriate free benzhydrylamine in 10 ml of C₆H₆. The mixture was kept at 20–25° for 12 hr and then filtered. The filtrate was treated with an excess of the appropriate amine and the mixture was refluxed for 15 hr. After cooling, C₆H₆ extract was filtered, washed twice with H₂O, and dried (MgSO₄), the solvent was evaporated, and the hydrochloride was prepared (EtOH-ether).

2,2'-Dialkylbenzhydryl Esters.—A soln of 0.01 mole of Cl-COCH₂Cl or BrCOCHBrCH₃ in 10 ml of C₆H₆ was added over a period of 0.5 hr to a cold, stirred soln of 2.12 g (0.01 mole) of di-*o*-tolylcarbinol¹⁰ and 0.89 ml (0.011 mole) of pyridine in 30 ml of C₆H₆. The mixture was stirred an addn 0.5 hr. It was filtered, the filtrate treated with an excess of the appropriate amine, and the mixture kept at 40–45° for 3 days. After cooling, it was treated as described in the above procedure.

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(10) H. H. Hatt, *J. Chem. Soc.*, 1631 (1929).

Molecular Orbital Conformation of Phenyl Choline Ether

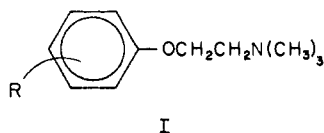
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Over the past decade a considerable amount of theoretical interest has been shown in the structural considerations of the nicotinic activity of a variety of molecules. Fukui studied a series of phenyl choline ethers, (I), using simple Hückel MO calculations.¹



The study revealed that the substituent groups influence frontier electron density at the O and the superdelocalizability at the ortho positions, and that these indices correlated with nicotinic activity. The suggestion was made that the O atom and the ortho position on the ring might be spatially related to the onium group in the phenyl choline ethers in the same

manner as the ether and CO oxygen atoms in the potent nicotinic agent, ACh (Figure 1). Thus an ortho ring position would play a role at the receptor comparable to the carbonyl O atom.

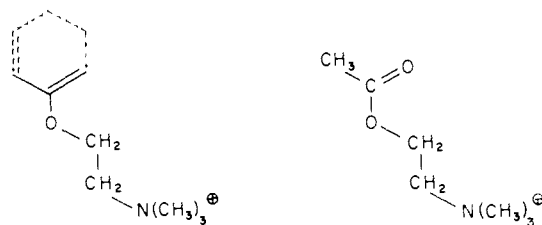


Figure 1.—Spatial relationship between phenyl choline ether and acetylcholine postulated by Fukui¹ and predicted from this study.

We have reported extended Hückel MO calculations on both ACh² and nicotine³ in which we concluded that both nicotine and ACh can assume preferred conformations in which the pyridine N and carbonyl O atoms, respectively, are of similar distance from the onium groups in each molecule (Figure 2). The role

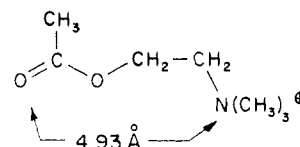
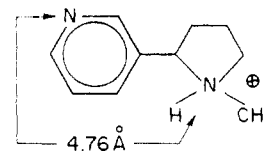


Figure 2.—Spatial relationship between nicotine and acetylcholine in their predicted preferred conformations.³

of the ether O in the nicotinic activity of ACh is not evident from this comparison, since there is no atom in nicotine that appears equivalent to the ether O at a receptor. The receptor equivalence of the pyridine N and the carbonyl O is certainly suggested from these studies.

In a recent study by Crow, *et al.*, a series of ring-substituted phenyl choline ethers was considered using simple Hückel theory.⁴ The relationships between calculated reactivity indices and nicotinic activity revealed correlations only with the energy of the highest occupied MO and the superdelocalizability at the ring ortho positions. These findings agree with those of Fukui¹ except for the lack of a correlation between an oxygen reactivity index and activity. The suggestion arising from these results was that the aromatic ring interacts with the receptor, probably at one of the ortho positions, to form a charge-transfer complex.

A basic problem still remains with the phenyl choline ethers if the ortho ring position is to be seriously considered as mimicking the carbonyl O of ACh at the

(2) L. B. Kier, *Mol. Pharmacol.*, **3**, 487 (1967).

(3) L. B. Kier, *ibid.*, **4**, 70 (1968).

(4) J. Crow, O. Wassermann, and W. Holland, *J. Med. Chem.*, **12**, 764 (1969).

* To whom correspondence should be addressed.

(1) F. Fukui, C. Nagata, and A. Imamura, *Science*, **132**, 87 (1960).